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CURRENT ASPECTS OF ANTIBIOTIC TREATMENT AND ENDOTOXIN RELEASE

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Abstract

Endotoxin is a major cause of sepsis and organ failure in humans. Antibiotics, which are administrated to treat these severe infections, may release Endotoxin from the bacterial wall and may harm the patient. Penicillin-binding protein (PBP) 2-specific antibiotics, e.g., imipenem were considered to release less amounts of free Endotoxin than PBP 3-specific antibiotics, e.g., Ceftazidime. This effect has been contributed to an increased bactericidal activity of PBP 2-specific antibiotics and consecutive change in morphology of pathogens, enabling phagocytosis. Recent in vitro studies, however, were unable to repeat these results. The antibiotic-induced Endotoxin release may change with the type of pathogen and dosing of the antibiotic. In animal studies Endotoxin release did not show a correlation to the bactericidal effect in all experiments. Antibiotic-induced Endotoxin release and outcome was different with regard to animal models, location of infection, strains, pharmacodynamics and dosage of antibiotics. Bacteriostatic antibiotics, e.g., lincomycin and clindamycin, were able to induce Endotoxin release. In some studies imipenem caused either similar release of Endotoxin compared to ceftazidime or more compared to ciprofloxacin. Chemically modified tetracycline or combination of antibiotics prevented an increased Endotoxin release. In patients with urosepsis controversial results were observed when imipenem was compared to ceftazidime. In clinical observational studies or post-hoc analysis of a randomized clinical trial a differential release of Endotoxin after imipenem and cephalosporins has been reported. In conclusion, antibiotic-induced Endotoxin release may be clinically relevant. However, there are many interfering factors in clinical studies, which need to be addressed properly when analyzing studies on antibiotic-induced Endotoxin release.

Key words

Antibiotic-induced endotoxin release, LPS

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ASPECTOS ACTUALES DEL TRATAMIENTO ANTIBIOTICO Y LA LIBERACION DE ENDOTOXINAS

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Resumen

La endotoxina es una de las causas principales de sepsis e insuficiencia multiorgánica en los seres humanos. Los antibióticos que se administran para tratar estas infecciones graves pueden liberar endotoxina de la pared bacteriana y afectar al paciente. Se consideraba que los antibióticos específicos de la proteína ligadora de penicilina (PLP) 2, por ejemplo, imipenem, liberaban menores cantidades de endotoxina libre que los antibióticos específicos de la PLP 3, como la ceftazidima. Este efecto contribuye al aumento de la actividad bactericida de los antibióticos específicos de la PLP 2, con los consiguientes cambios en la morfología de los patógenos, lo que posibilita la fagocitosis. Sin embargo, recientes estudios *in vitro* no pudieron repetir estos resultados. La liberación de endotoxina inducida por antibióticos puede cambiar con el tipo de patógeno y la dosificación del antibiótico. En estudios con animales, la liberación de endotoxina no se correlacionó con el efecto bactericida en todos los experimentos. La liberación de endotoxina inducida por antibióticos, así como los resultados, fue diferente según los modelos con animales, la localización de la infección, las cepas, la farmacodinamia y la dosificación del antibiótico. Los antibióticos bacteriostáticos, como lincomicina y clindamicina, indujeron la liberación de endotoxina. En algunos estudios la liberación de endotoxina inducida por imipenem fue similar a la causada por ceftazidima y mayor que la inducida por ciprofloxacina. Las tetraciclinas modificadas químicamente y la combinación de antibióticos evitaron el aumento en la liberación de endotoxinas. En pacientes con urosepsis se observaron resultados controvertidos al comparar imipenem con ceftazidima. En estudios clínicos de observación o en los análisis *post hoc* de ensayos clínicos aleatorizados se informaron diferencias en la liberación de endotoxina luego de la administración de imipenem y cefalosporinas. En conclusión, la liberación de endotoxina inducida por antibióticos podría ser clínicamente relevante. No obstante, en los estudios clínicos pueden interferir muchos factores que deben ser abordados debidamente cuando se analizan los estudios acerca de este tema.

Palabras clave

Liberación de endotoxina inducida por antibióticos, lipopolisacáridos

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CURRENT ASPECTS OF ANTIBIOTIC TREATMENT AND ENDOTOXIN RELEASE

Endotoxin is one of the major pathogenic causes of gram-negative sepsis in humans.¹ The clinical significance of endotoxin and its association with mortality has been demonstrated in several studies.²⁻⁶ Recent reviews discussed drug-induced, e.g., antibiotic, fever as possible cause of fever of unknown origin.^{7,8} Since Jackson and Kropp (1992) demonstrated that Penicillin-binding Protein (PBP) 2-specific antibiotics (Imipenem) may release less free endotoxin than PBP 3-specific antibiotics (ceftazidime) which has been associated with a filamentation of the pathogen, it has been assumed that antibiotics may have a different potential to release endotoxin and to influence the outcome of septic patients.⁹ However, other bacterial wall compounds, Lipoteichoic acid (LTA), may be involved in the host response after antibiotic treatment.¹⁰

In vitro studies

Most studies were performed in vitro with different pathogens and antibiotics. The effect of penicillin has been investigated in *Neisseria meningitis*, *Streptococcus* group A, *Staphylococcus aureus*, *Meningococcus*, and *Streptococcus faecium*. In most trials penicillin released lipopolysaccharide (LPS), LTA, IL-1 and TNF-alpha. (table 1).

Table 1: Effects of penicillin on in-vitro endotoxin/cytokine/protein release

Year	Author	Pathogen or cell	Endotoxin/cytokine/protein release
1980	Anderssen ¹¹	N meningitis	LPS+
1981	Kessler ¹²	S group A	LTA, sLTA +
1985	Gold ¹³	S faecium	IL-1 +
1986	Nealon ¹⁴	S group A, S aureus	LTA +
1987	Kiryama ¹⁵	S aureus	LTA +
1991	Mellado ¹⁶	N meningitis	LPS+
1992	Iino ¹⁷	LPS	TNF +
1997	Prins ¹⁸	Meningococcus	LPS-

LPS Lipopolysaccharide; LTA Lipoteichoic acid; IL-1 Interleukin-1; TNF-alpha Tumor Necrosis Factor alpha

The effect of aminoglycoside therapy on endotoxin release has been studied mainly in *E. coli*, *K. pneumoniae*, *P. aeruginosa*, *H. influenzae*, *E. cloacae*, *S. aureus*, *Salmonella* spp., and *Neisseria* spp. Depending on pathogen gentamicin was able to increase or decrease endotoxin release or it showed no effect when compared to controls. Most studies reported a decrease in TNF-alpha release after treatment with an aminoglycoside. Tobramycin lead to a decreased release of LPS and amikacin induced a decrease in LPS and TNF release (table 2).

Table 2: Effects of amikacin/gentamicin on in-vitro endotoxin/cytokine/protein release

Year	Author	Aminoglycoside	Pathogen or cell	Endotoxin/cytokine/protein release
1975	Rusmin ¹⁹	G	E coli, K pneumoniae, P aeruginosa	LPS+
1985	Shenep ²⁰	G	E coli	LPS+
1986	Cohen ²¹	G	E coli	LPS-
1989	Stokes ²²	G		TNF +
1991	Simon ²²	A	E coli, THP cell	TNF-
1992	Binger ²³	A	H influenzae	LPS-
1992	Van den Berg ²⁴	G	E coli	LPS+
1993	Eng ²⁵	G	E coli K pneumoniae E cloacae P aeruginosa S aureus	LPS- LPS- No effect No effect No Endotoxin
1993	Evans ²⁶	G	E coli	LPS+
1994	Crosby ²⁷	T	E coli, E cloacae	LPS-
1995	Prins ²⁸	G	E coli	TNF-
1996	Lamp ²⁹	A	E coli, P aeruginosa	LPS-
1998	Van Langevelde ³⁰	G	S aureus	LTA, PG+
1998	Trautmann ³¹	T	E coli	LPS-
2000	Sjolin ³³	T	E coli	LPS-
2001	Xu ³⁴	A	P aeruginosa, E coli	LPS-
2002	Bentley ³⁵	A	E coli, whole blood	LPS-, TNF-
2002	Krehmeier ³⁶	G	PBMC	TNF-
2003	Tsumura ³⁷	G	E coli	LPS+
2003	Goscinski ³⁸	T	E coli, Klebsiella, Salmonella, Neisseria	LPS-

LPS Lipopolysaccharide; LTA Lipoteichoic acid; IL-1 Interleukin-1; TNF-alpha Tumor Necrosis Factor alpha
G gentamicin; A amikacin; T tobramycin

In most studies imipenem, the compound which was studied best was able to decrease endotoxin release or TNF-alpha/IL-6 release. However, in recent trials^{35,39} imipenem either increased endotoxin/TNF-alpha release or had no effect (table 3).

Table 3: Effects of imipenem on in-vitro endotoxin/cytokine/protein release

Year	Author	Pathogen or cell	Endotoxin/cytokine/protein release
1991	Simon ⁷³	E coli, THP cell	TNF-
1991	Dofferhoff ⁸⁰	E coli	LPS-
1992	Jackson ⁹	P aeruginosa	LPS-
1993	Arditi ⁴¹	H influenzae	TNF-
1993	Evans ¹⁶	E coli	LPS+
1993	Dofferhoff ⁸²	E coli	TNF 4h similar, 24h -
1993	Eng ⁷³	E coli, K pneumoniae, E cloacae, P aeruginosa S aureus	LPS- LPS- LPS- LPS- No endotoxin
1995	Prins ⁷⁸	E coli, whole blood	LPS-, TNF-, IL-6-
1996	Yokochi ⁴³	P aeruginosa	LPS-
1996	Inoue ⁴⁴	E coli, S aureus, E cloacae, C freundii, P aeruginosa, S maltophilia	LPS-
1996	Lamp ⁷⁹	E coli, P aeruginosa	LPS-
1997	Narita ⁴³	P aeruginosa	LPS-
1997	Arditi ⁴⁶	E coli	IL-6-
1997	Takahashi ⁴⁷	E coli	LPS-
1998	Trautmann ³¹	E coli	LPS-
1998	Van Langevelde ⁴⁸	S typhi	No difference
1998	Hori ⁴⁹	E coli, S marcescens, K pneumoniae, P aeruginosa, P mirabilis, P vulgaris	LPS- (exception P aeruginosa)
1998	Trautmann ³⁰	P aeruginosa	LPS-
1999	Yamaguchi ³¹	P aeruginosa	LPS-
1999	Trautmann ³⁷	E coli, Monocyte	LPS-, TNF-
1999	Hori ³³	P aeruginosa	LPS-
2000	Cui ¹⁴	E coli, S aureus	TNF+ release in co-culture with S aureus
2001	Xu ²⁴	P aeruginosa, E coli	LPS-
2002	Bentley ³³	E coli, whole blood	LPS+, TNF+
2003	Tsujii ³⁷	P aeruginosa	LPS no difference

LPS Lipopolysaccharide; IL-1 Interleukin-1; TNF-alpha Tumor Necrosis Factor alpha; IL-6 Interleukin-6

Ceftazidime is the compound used in most in-vitro studies on endotoxin release. In the majority of studies endotoxin or proinflammatory cytokines, e.g., TNF-alpha, IL-6 and IL-1 were increased when ceftazidime was used for treatment. However, ceftazidime decreased LPS-release from *P. aeruginosa*²⁵ or did not show a difference compared to untreated controls^{37,49} (table 4).

Table 4: Effects of ceftazidime on in-vitro endotoxin/cytokine/protein release

Year	Author	Pathogen or cell	Endotoxin/cytokine/protein release
1991	Simon ²³	E coli, THP cell	TNF+
1991	Dofferhoff ²⁰	E coli	LPS+
1992	Jackson ⁹	P aeruginosa	LPS+
1993	Evans ¹⁶	E coli	LPS+
1993	Dofferhoff ²¹	E coli	TNF+
1993	Eng ²¹	E coli, K pneumoniae, E cloacae, P aeruginosa S aureus	LPS+ or no effect LPS+ or no effect LPS+ or no effect LPS- No Endotoxin
1994	Bucklin ¹³	E coli	LPS+
1994	Leeson ¹⁶	Monocytes E coli	TNF+
1995	Prins ²³	E coli, whole blood	LPS+, TNF+, IL-6+
1996	Inoue ⁴⁴	E coli, S aureus, E cloacae, C freundii, P aeruginosa, S maltophilia	LPS+
1996	Lamp ¹⁹	E coli, p aeruginosa	LPS+
1997	Narita ⁴³	P aeruginosa	LPS+
1998	Trautmann ³¹	E coli	LPS+
1998	Van Langevelde ⁴³	S typhi	LPS no difference
1998	Hori ⁴⁹	E coli, S marcescens, K pneumoniae, P aeruginosa, P vulgaris, P mirabilis	LPS+ (exception P aeruginosa)
1998	Trautmann ³⁰	P aeruginosa	LPS+
1999	Kishi ¹⁷	E coli	LPS+, TNF+, IL-1+
1999	Yamaguchi ³¹	P aeruginosa	LPS+
1999	Trautmann ³⁷	E coli, Monocyte	LPS+, TNF+
2000	Hori ³³	E coli	LPS+
2001	Xu ²⁴	P aeruginosa, e coli	LPS+
2003	Tsuji ¹⁷	P aeruginosa	LPS no difference
2003	Liang ²⁹	P aeruginosa, RAW cells	LPS+, TNF+

LPS Lipopolysaccharide; IL-1 Interleukin-1; TNF-alpha Tumor Necrosis Factor alpha

The best studied quinolones are ciprofloxacin and ofloxacin. In the some recent studies ciprofloxacin or ofloxacin were responsible for a decrease in endotoxin release or TNF-alpha/IL-6 release. However, some investigators reported an increase in endotoxin and/or proinflammatory cytokine release^{17,21,24,28,29} (table 5).

Table 5: Effects of ciprofloxacin/ofloxacin on in-vitro endotoxin/cytokine/protein release

Year	Author	Quinolone	Pathogen or cell	Endotoxin/cytokine/protein release
1986	Cohen ²¹	C	E coli	LPS+
1991	Simon ²³	C	E coli, THP cell	TNF intermediate
1992	Iino ¹⁷	O	Monocytes	TNF+
1992	Van den Berg ²⁴	C	E coli	LPS+
1993	Eng ²⁵	C O	E coli, K pneumoniae, E cloacae, P aeruginosa S aureus	LPS- LPS- LPS- LPS+ or no effect No endotoxin
1994	Crosby ²⁷	C	E coli, E cloacae	LPS+
1995	Prins ²⁸	C	E coli, whole blood	LPS+, TNF+, IL-6+
1996	Lamp ²⁹	O	E coli, P aerugi- nosa	LPS-
1998	Trautmann ³¹	C	E coli	LPS-
1999	Trautmann ³²	C	E coli	LPS-
2002	Krehmeier ³⁶	O C	PBMC PBMC	TNF-, IL-6- TNF-

LPS Lipopolysaccharide; IL-1 Interleukin-1; TNF-alpha Tumor Necrosis Factor alpha; IL-6 Interleukin-6
C ciprofloxacin; O ofloxacin

Clindamycin and erythromycin were able to reduce the release of LPS and pro-inflammatory cytokines in gram-negative pathogens, but also lipoteichoic acid (LTA) and proinflammatory cytokines in gram-positive pathogens in most studies (table 6).

Table 6: Effects of clindamycin/erythromycin on in-vitro endotoxin/cytokine/protein release

Year	Author	Antibiotic	Pathogen or cell	Endotoxin/cytokine/protein release
1986	Nealon ¹⁴	C	S group A, S aureus	LTA S aureus -
1992	Iino ¹⁷	E	Monocytes	TNF-
1995	Khair ⁶⁰	E	H influenzae, HBEC	IL-6 -, IL-8 - sICAM-1 -
1998	Van Langevelde ³⁰	E C	S aureus, whole blood	LTA -, PG unchanged, TNF -, IL-10 -
1999	Kishi ⁵⁷	C	E coli	LPS -, IL-1 -, TNF -
2000	Orman ⁶¹	C	S pneumoniae, murine macro- phages	iNOS -, TNF-
2000	Hori ⁵⁸	C	E coli	LPS-
2003	Gerber ⁶²	C	S pneumoniae	LTA -

E erythromycin, C clindamycin

HBEC human bronchial epithelial cells

LPS Lipopolysaccharide; LTA Lipoteichoic acid; IL-1 Interleukin-1; TNF-alpha Tumor Necrosis Factor alpha;
IL-6 Interleukin-6; IL-8 Interleukin-8; IL-10 Interleukin-10; PG Peptidoglycan; iNOS inducible Nitric Oxide
Synthase; sICAM-1 soluble Intercellular-Adhesion-Molecule 1

Animal studies

Endotoxin liberating strains were considered to be more virulent.¹¹ An increase in endotoxin plasma levels after antibiotic administration has been observed by Shenep in 1984.⁶³ In 1986 Walterspiel demonstrated that subinhibitory doses of Polymyxin B modulate the lethal effects of LPS.⁶⁴ Shenep (1985) postulated that the LPS release is dependent on the class of antibiotics and

does not correlate with the rate of bacterial killing.²⁰ Johnston reported in 1984 that the level of endotoxemia was higher in antibiotic-treated survivors than in animals dying without antibiotics.⁶⁵

Pre-treatment in animal studies

Selective antibiotic decontamination resulted in increased LPS levels.^{66,67} Pre-treatment with agents that altered gut contents reduced endotoxemia and mortality.⁶⁸ Elevated TNF-release may be associated with enhanced hemodynamic response but not with increased mortality.⁶⁹ Pre-treatment with erythromycin may have beneficial effects in *Candida albicans* infection.⁷⁰

Mode of application, location of infection

LPS release is not only influenced by the mode of action of the antibiotic class, but also by dosage, pharmacodynamics.⁷¹ Topically applied imipenem demonstrated a profound bactericidal effect, but also increased LPS/TNF-alpha release.⁷² Meropenem may increase LPS-release and mortality, whereas imipenem does the opposite.⁴⁵ The morphological changes induced by imipenem treatment may help phagocytosis by peritoneal cells.⁷³ In a burn injury model imipenem released less LPS but there was no relationship to a bactericidal effect.⁷⁴ Imipenem induced highest levels of LPS in *B. fragilis* and *Fusobacterium* spp. infection.⁷⁵ LPS release may not be influenced by the bactericidal effect: imipenem and ceftazidime had a similar bactericidal power in a rat sepsis model, but there was less LPS release after imipenem treatment.⁷⁶ In 2003, Tsuji et al. demonstrated that imipenem, doripenem, meropenem and ceftazidime induced similar serum levels of LPS.³⁷ Treatment of animals with intra-abdominal infection resulted in increased survival, decreased plasma and peritoneal fluid levels of cytokines. LPS release was higher after imipenem than after ciprofloxacin treatment.⁷⁷

Meningitis

In meningitis cefotaxime, ceftazidime, meropenem and gentamicin induced endotoxin release. However, untreated animals showed higher endotoxin levels.⁷⁸ In *H. influenzae* infection of the middle ear no LPS has been observed after treatment with ceftriaxone.⁷⁹ In rabbit *S. pneumoniae* meningitis initiation of therapy with clindamycin (protein-synthesis inhibition) and continuation with a combination of ceftriaxone (beta-Lactam antibiotic) decreased neuronal injury.⁶² This has been supported by a further study by Bottcher (2004).⁸⁰

Bacteriostatic and combination antibiotic treatment

Bacteriostatic antibiotics (lincomycin and clindamycin) induced LPS release compared to no treatment.⁵⁸ Doxycycline exerted its protective effect by inhibiting nitrate production in a BALB mouse LPS model.⁸¹ Recent studies investigated the effect of other compounds, matrix metalloproteinase inhibitors – chemically modified tetracycline, which prevented acute lung injury after cardiopulmonary bypass.⁸² These compounds can significantly preserve cardiac mechanical function during septic shock.⁸³ The combination treatment with other endotoxin-neutralizing compounds, e.g., lipopolyamines⁸⁴ or Bactericidal-Permeability-increasing-Protein (BPI21)⁸⁵ may help to prevent the cephalosporin-induced increase in LPS release and improve survival. Mice challenged by *E. coli* showed a change in LPS-induced cytokine production and improved survival when pre-treated by clindamycin.⁸⁶ Antibiotics resulted in a shift of LD50 of approximately 500-fold in *E. coli* mouse infection model.⁸⁷ In 2003, Tsumura demonstrated in a rabbit *E. coli* model that flomoxef and gentamicin reduced in-vivo plasma LPS, TNF-alpha and blood bacterial counts to comparable levels. LPS release may not be a problem if appropriate antimicrobial agents were used.³⁷

Clinical studies
In 1983 Teklu reported on the beneficial effects of mepizolinol, an opioid antagonist with agonist properties, which diminished the Jarisch-Herxheimer reaction after tetracycline treatment in a randomized study in patients with louse-borne fever.⁸⁸

Meningitis
In *H. influenzae* meningitis ceftriaxone treatment leads to a release of free LPS with an associated inflammatory response.⁸⁹ Intraventricular applied gentamicin may have caused release of LPS resulting in increased IL-1 concentrations and poor outcome of patients with *coliform meningitis*.⁹⁰

Urosepsis
Several studies were performed in patients with urosepsis. Imipenem caused less LPS release, serum and urine cytokine levels when compared to cefazidime.⁹¹ An increased release of endotoxin and TNF was observed in patients with acute pyelonephritis. It might be responsible for the persistence of fever despite negative blood cultures.⁹² In a RCT in patients with gram-negative urosepsis no difference was found in plasma endotoxin, proinflammatory cytokines within the first 8 hours after antibiotic treatment with imipenem or cefazidime.⁹³

Trauma and surgical intensive care patients
In trauma patients antibiotics that are associated with a greater release of endotoxin and TNF (aztreonam, cefazidime, and cefotaxime) are associated with greater release of endotoxin and TNF, and a greater mortality.⁴ In surgical intensive care patients a significant increase of endotoxin plasma levels were observed after treatment with cefotaxime and ceftriaxone when compared to ciprofloxacin, tobramycin, and imipenem.⁹⁴ Patients undergoing hepatic resection showed no elevation in peripheral blood endotoxin levels after treatment with cefmetazole, latamoxef, flomoxef, cefazolin, cefoperazone, ceftioam.

The authors concluded that the endotoxin has been scavenged from the blood.⁹⁵ In 2001 Byl et al. observed a similar effect on endotoxin and cytokine release in gram-negative infections when treated with cefazidime and imipenem.⁹⁶ Maskin reported high plasma concentrations of LPS, IL-6, and TNF-alpha after treatment with cefazidime and imipenem in septic patients. However, TNF-alpha plasma levels were significantly less 4 hours after imipenem administration.⁹⁷ In aortic aneurysm repair, a clinical model for reperfusion-injury and associated endotoxin release, oral ofloxacin prophylaxis lead to alteration of endotoxin neutralizing capacity and IL-6 plasma levels, but had no effect on endotoxin plasma levels and other inflammatory mediators.⁹⁸ In melioidosis caused by *Burkholderia pseudomallei* treatment with imipenem reduced the plasma endotoxin release without affecting survival.⁹⁹ In burn patients' endotoxin and TNF release increased 2h after cefoperazone treatment when compared to imipenem¹⁰⁰ (table 7).

Table 7: Clinical studies in antibiotic-induced endotoxin release

Year	Author	Type of study	Type of infection	Antibiotics
1983	Teklu	RCT	Louse-borne relapsing fever	Tetracycline
1989	Ardifi	Prospective observation	H influenzae meningitis	Ceftaxone
1989	Mustafa	RCT	Coliform meningitis	Gentamicin
1995	Prins	RCT	Urosepsis	Imipenem Ceftazidime
1995	Mock	Post-hoc analysis	Trauma patients	Aztreonam Ceftazidime Cefotaxime
1996	Holzheimer	Prospective observational	Surgical intensive care	Ciprofloxacin Cefotaxime Tobramycin Ceftriaxone Vancomycin Imipenem
1999	Ishikawa	Prospective observation	Hepatic resection	Cefmetazole Latamoxef Flomoxef Cefazolin Cefoperazone Cefotiam
1999	Giamarellou-Bourboulis	Prospective observation	Acute pyelonephritis	Cefuroxime
2000	Simpson	RCT	Melioidosis caused by Burkholderia pseudomallei	Ceftazidime Imipenem
2000	Luchi	RCT	Gram-negative urosepsis	Ceftazidime Imipenem
2001	Byl	RCT	Gram-negative infection	Ceftazidime Imipenem
2001	Jaber ^{1b1}	Prospective observation		Gentamicin
2002	Maskin	RCT	Sepsis	Ceftazidime Imipenem
2003	Holzheimer	RCT	AAA repair	Ofloxacin Cefotiam
2004	Wang	RCT	Burn patients with gram-negative infection	Imipenem Cefoperazone

RCT randomized controlled trial

Summary

Endotoxin is a major cause of sepsis and organ failure in humans. Antibiotics are administered to treat these severe infections. However, antibiotics may also do harm if not applied properly or may release Endotoxin from the bacterial wall, which may harm the patient. Penicillin-binding protein (PBP) 2-specific antibiotics, e.g., imipenem were considered to release less amounts of free Endotoxin than PBP 3-specific antibiotics, e.g., Ceftazidime. This effect has been contributed to an increased bactericidal activity of PBP 2-specific antibiotics and consecutive change in morphology of pathogens, enabling phagocytosis. Recent in vitro studies, however, were unable to repeat that result. The antibiotic induced Endotoxin release may be dependent on the strain and dosing of the antibiotic. In animal studies Endotoxin release was not influenced by the bactericidal effect. Antibiotic-induced Endotoxin release and outcome was different with regard to animal models, location of infection, strains, pharmacodynamics and dosage of antibiotics. Bacteriostatic antibiotics, e.g., lincomycin and clindamycin, were able to induce Endotoxin release. Imipenem caused either similar release of Endotoxin compared to Ceftazidime or more compared to ciprofloxacin in some studies.

Chemically modified tetracycline or combination of antibiotics prevented an increased Endotoxin release. In patients with urosepsis controversial results were observed when imipenem was compared to Ceftazidime. In clinical observational studies or post-hoc analysis a differential release of Endotoxin after imipenem and cephalosporins has been reported. In conclusion, antibiotic-induced Endotoxin release may be clinically relevant. Analysis of clinical studies which failed to demonstrate the effect of antibiotic-induced Endotoxin release may help to plan further studies.

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